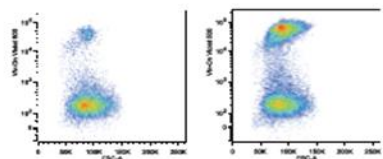


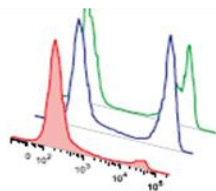
## SELECTION GUIDE

Fixable Viability Dye	Excitation source (nm)	Emission (nm)
VIV-ON VIOLET 500 FVD	405	515
VIV-ON BLUE 520 FVD	488	523
VIV-ON RED 660 FVD	633	660
VIV-ON RED 780 FVD	633	780

## FIGURES



EHEB cells were untreated (left) or treated 10 min at 60°C (right), then stained with Viv-On Violet 500 FVD and fixed. Total cells were used for analysis.



■ EHEB cells untreated  
 ■ EHEB cells, treated and stained  
 ■ EHEB cells, treated, stained and fixed

Downloads: <http://www.cyanagen.com/downloads/product-manuals#family-2>

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## VIV-ON FIXABLE VIABILITY DYES FOR FLOW CYTOMETRY

### TECHNICAL DESCRIPTION

**VIV-ON Fixable Viability Dyes** are used to distinguish live cells from dead cells, based on cell membrane integrity.

VIV-ON Fixable Viability Dyes are amine reactive and membrane impermeable.

VIV-ON Fixable Viability Dyes are available for the 405-, 488- and 633 nm laser lines, with detection in the common green, red and infrared channels.

### COMPONENTS

Vial A: VIV-ON Dye

Vial B: VIV-ON FVD – DMSO, Cod. FV160,200

### STORAGE CONDITIONS

Protect from light. Store at -20°C with desiccant.

## VIV-ON FIXABLE VIABILITY DYES FOR FLOW CYTOMETRY

### FEATURES

- High brightness for optimal differentiation between live and dead cells
- Ready-to-use kit
- Unlike 7-AAD and PI, labelled cells can be fixed, permeabilized, washed and stained
- May be used for any cell species

### VIV-ON Fixable Viability Dye solution

Before the first use, add 100 µL of VIV-ON FVD – DMSO to the vial VIV-ON – Dye.  
Store the solution at -20°C.

## VIV-ON FIXABLE VIABILITY DYES FOR FLOW CYTOMETRY

### GENERAL PROTOCOL ASSAY

1. Prepare cells as desired.
2. Wash cells twice in azide-free and protein-free phosphate buffer saline (PBS).
3. Resuspend cells at 1-10 x10<sup>6</sup> cells / mL in azide-free and protein-free phosphate buffer saline (PBS).  
**Note:** For consistent staining it is not recommended to stain in less than 0.5 mL.
4. Stain cells by adding 1 µL of VIV-ON Fixable Viability Dye solution per 1 mL of cells and mix by vortexing.
5. Incubate cells for 30 minutes at 2-8 °C. Protect from light.
6. Wash cells twice in phosphate buffer saline (PBS) or any appropriate flow cytometry buffer.
7. Proceed with experiment, as desired.